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Datasheet/User Guide

Mouse B7H1/PD-L1 ELISA Kit

Catalog Number: E92-031

For Serum, Plasma, Cell Culture Supernatants

Introduction

The Mouse B7-H1/PD-L1 ELISA (Enzyme-Linked Immunosorbent Assay) kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of mouse B7-H1 in serum (mouse B7-H1 concentration is low in normal serum/plasma, and may not be detectable in this assay), plasma and cell culture supernatants. This assay employs an antibody specific for mouse B7-H1 coated on a 96-well plate. Standards and samples are pipetted into the wells and B7-H1 present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated anti-mouse B7-H1 antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of B7-H1 bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

Storage/Shelf Life

2 - 8°C/6 months from date of receipt

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Reagents

Component	Size / Description	Storage/Stability After Preparation
Mouse B7-H1/PD-L1 Microplate	96 wells (12 x 8) coated with anti-Mouse B7-H1/PD-L1	1 month at 4ºC
Mouse B7-H1/PD-L1 Standard Protein	2 vials Mouse B7-H1/PD-L1 1 vial is enough to run each standard in duplicate	1 week at -80ºC
Mouse B7-H1/PD-L1 Detection Antibody	2 vials biotinylated anti-Mouse B7-H1/PD-L1; Each vial is enough to assay half the microplate	5 days at 4ºC
Wash Buffer	25 ml 20X concentrated solution	1 month at 4ºC
HRP-Streptavidin	200 µL 1000X concentrated HRP-conjugated Streptavidin	Do not store and reuse
TMB One-Step Substrate Reagent	12 ml 3,3,5,5'-tetramethylbenzidine (TMB) in buffer solution	n/a
Stop Solution	8 ml 0.2 M sulfuric acid	n/a
Assay Diluent D	15 ml 5X concentrated diluent buffer	n/a
Assay Diluent B	15 ml 5X concentrated diluent buffer	1 month at 4ºC

Additional Materials Required (but not provided)

- Microplate reader capable of measuring absorbance at 450 nm.
- ELISA plate shaker or platform rocker.
- Precision pipettes to deliver 2 µl to 1 ml volumes.
- Adjustable 1-25 ml pipettes for reagent preparation.
- 100 ml and 1 liter graduated cylinders.
- Absorbent paper.
- Distilled or deionized water.
- Log-log graph paper or computer and software for ELISA data analysis.
- Tubes to prepare standard or sample dilutions.

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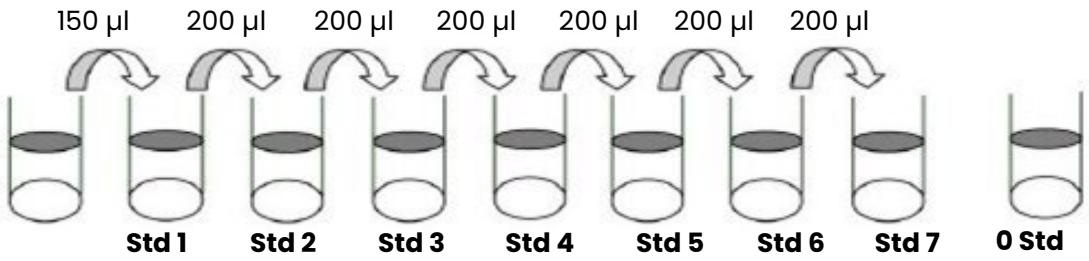
Reagent Preparation

1. Bring all reagents and samples to room temperature before use.
2. Assay Diluent D and Assay Diluent B should be diluted 5-fold with deionized or distilled water before use.
3. Sample dilution: 1X Assay Diluent D should be used for dilution of serum, plasma, and cell culture supernatant samples. The suggested dilution for normal serum/plasma is 2-fold.

   Note: Levels of B7H1/PD-L1 may vary between different samples. Optimal dilution factors for each sample must be determined by the investigator.

4. Preparation of standard: Briefly spin a vial of standard protein. Add 600 µL Assay Diluent D (1X) (should be diluted 5-fold with deionized or distilled water before use) into the Standard Protein vial to prepare a 50 ng/mL standard solution. Dissolve the powder thoroughly by a gentle mix. Add 150 µL of B7-H1 standard solution from the vial of Standard Protein, into a tube with 350 µL Assay Diluent (1X) to prepare a 15 ng/mL standard solution. Pipette 300 µL Assay Diluent D (1X) into each tube. Use the 15 ng/mL standard solution to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. Assay Diluent D (1X) serves as the zero standard (0 ng/mL).



Diluent volume	Std + 600µl	350 µl	300 µl	300 µl	300 µl	300 µl	300 µl	300 µl	300 µl
Concentration	50 ng/ml	15 ng/ml	6 ng/ml	2.4 ng/ml	0.96 ng/ml	0.384 ng/ml	0.154 ng/ml	0.061 ng/ml	0 ng/ml

5. If the Wash Buffer (20X) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 25 ml of 20X Wash Buffer into 475 ml deionized or distilled water to yield 500 ml of 1X Wash Buffer.
6. Briefly spin the Detection Antibody vial before use. Add 100 µl of 1X Assay Diluent B into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4ºC for 5 days). The detection antibody concentrate should be diluted 80-fold with 1X Assay Diluent B and used in step 5 Assay Procedure.
7. Briefly spin the HRP-Streptavidin concentrate vial and pipette up and down to mix gently before use, as precipitates may form during storage. HRP-Streptavidin concentrate should be diluted 1000-fold with 1X Assay Diluent B. For example: Add 10 µl of HRP-Streptavidin concentrate into a tube with 10 ml 1X Assay Diluent B to prepare a final 1000-fold diluted solution. Mix well. DO NOT STORE THE DILUTED SOLUTION FOR NEXT DAY USE.

Assay Procedure

1. Bring all reagents and samples to room temperature before use. It is recommended that all standards and samples be run at least in duplicate.
2. Label removable 8-well strips as appropriate for your experiment.
3. Add 100 µl of each standard (see Reagent Preparation step 4) and sample into appropriate wells. Cover wells and incubate for 2.5 hours at room temperature with gentle shaking.
4. Discard the solution and wash 4 times with 1X Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel Pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 µl of 1X prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
6. Discard the solution. Repeat the wash as in step 4.
7. Add 100 µl of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
8. Discard the solution. Repeat the wash as in step 4.
9. Add 100 µl of TMB One-Step Substrate Reagent to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
10. Add 50 µl of Stop Solution to each well. Read at 450 nm immediately.

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Assay Procedure Summary

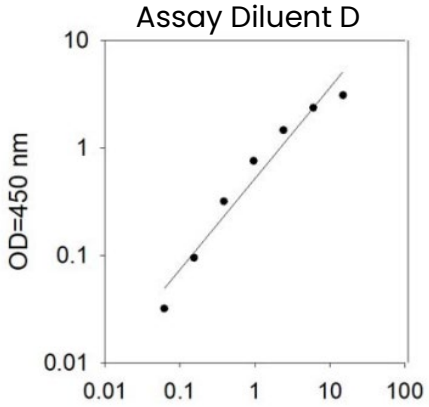
1. Prepare all reagents, samples and standards as instructed.
2. Add 100 µl standard or sample to each well. Incubate 2.5 hours at room temperature.
3. Add 100 µl prepared biotin antibody to each well. Incubate 1 hour at room temperature.
4. Add 100 µl prepared Streptavidin solution. Incubate 45 minutes at room temperature.
5. Add 100 µl TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
6. Add 50 µl Stop Solution to each well. Read at 450 nm immediately.

Calculation of Results

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigmaplot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

Typical Data

The typical standard curve was generated using the Mouse B7H1/PD-L1 ELISA kit protocol. The standard curves are for demonstration only. A standard curve must be generated for each assay.



Sensitivity

The minimum detectable dose of Mouse B7H1/PD-L1 was determined to be 0.055 ng/ml.

Minimum detectable dose is defined as the analyte concentration resulting in an absorbance that is 2 standard deviations higher than that of the blank (diluent buffer.)

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Spiking & Recovery

Recovery was determined by spiking various levels of Mouse B7H1/PD-L1 into the sample types listed below. Mean recoveries are as follows:

Sample Type	Average % Recovery	Range (%)
Serum	124.1	109-140
Plasma	123.3	108-137
Cell culture media	98.96	86-117

Linearity

Sample Type	Serum	Plasma	Cell culture media
1:2 dilution Avg % of Expected Range (%)	 120 106-130	 119 113-124	 123.7 113-135
1:4 dilution Avg % of Expected Range (%)	 113.3 112-116	 119.5 116-125	 116.9 101-133

Reproducibility

Intra-Assay CV%: <10%

Inter-Assay CV%: <12%

Specificity

This ELISA kit detects Mouse B7-H1/PD-L1. Other species not determined.

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Troubleshooting Guide

Problem	Potential Cause	Solution
Poor Standard Curve	◦Inaccurate pipetting ◦Improper standard dilution	◦Check pipettes ◦Briefly centrifuge the standard protein and dissolve the powder thoroughly by gently mixing
Low Signal	◦Improper preparation of standard and/or biotinylated antibody	◦Briefly spin down vials before opening
	◦Too brief incubation times	◦Ensure sufficient incubation times are followed. Standard/sample incubation step may be done overnight at 4ºC with gentle shaking (may increase overall signal, including background)
	◦Inadequate reagent volumes or improper dilution	◦Check pipettes and ensure correct preparation
Large CV	◦Inaccurate pipetting ◦Air bubbles in wells	◦Check pipettes ◦Carefully remove air bubbles
High Background	◦Plate is insufficiently washed	◦Review protocol for proper wash steps. If using a plate washer, ensure all ports are unobstructed.
	◦Contaminated wash buffer	◦Make fresh wash buffer
Low Sensitivity	◦Improper storage of the kit	◦Store the standard at -70ºC after reconstitution, others at 2-8 ºC. Keep substrate solution protected from light
	◦Stop Solution issue	◦Add stop solution to each well before reading plate

Warranty

Products are warranted by Bethyl Laboratories, Inc. to meet stated product specifications and to conform to label descriptions when used, handled and stored according to instructions. Unless otherwise stated, this warranty is limited to six months from date of receipt. Bethyl Laboratories sole liability for the product is limited to replacement of the product or refund of the purchase price. Bethyl Laboratories products are supplied for research applications. They are not intended for medicinal, diagnostic or therapeutic use. The products may not be resold, modified for resale or used to manufacture commercial products without prior written approval from Bethyl Laboratories, Inc. For more information, go to our website: https://www.fortislife.com/sales-terms